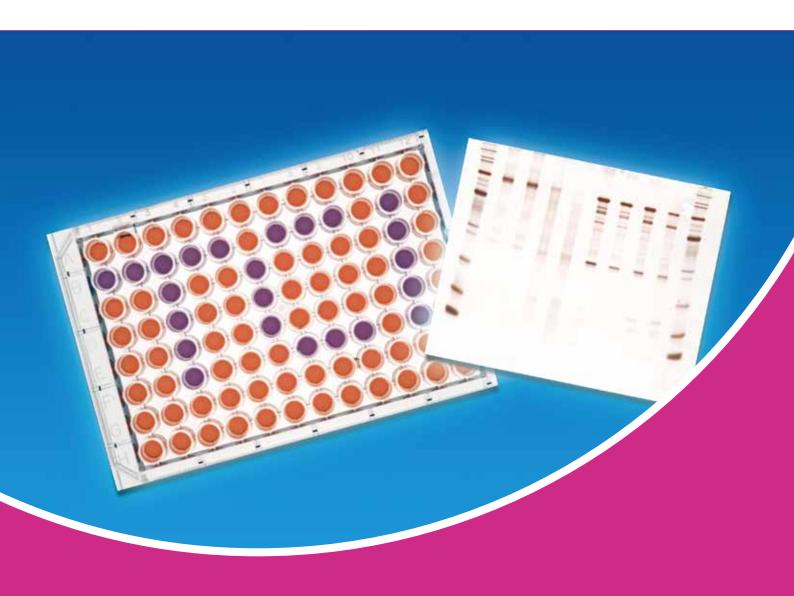




# **Protein and Biological Sample Preparation Reagents**



### Surfactants/Detergents

Surfactants are amphiphilic compounds containing both hydrophobic and hydrophilic groups and thereby are soluble in both organic solvents and water. Owing to the characteristic behavior of surfactants to orient at surfaces and form micelles by reducing the surface tension, they play an important role in many practical applications including the solubilization of membrane proteins, the decrease of the nonspecific adsorption of the material to the container surface etc. in the area of biochemistry. Surfactants are classified as ionic or nonionic depending on the formal charge on their hydrophilic head. Ionic surfactants can further be cationic, anionic or zwitter-ionic on the basis of the type of the charge present. The selection of surfactants in different fields is decided according to their particular usage.

### **Anionic Surfactants**

Lithium Dodecyl Sulfate (=LDS)	5g / 25g [L0254]
Sodium Dodecyl Sulfate (=SDS)	25g / 500g <b>[S0588]</b>
Tris Dodecyl Sulfate	250mg / 1g [T3071]
Sodium Deoxycholate	25g [D1820]
Sodium Cholate	5g/25g <mark>[S0596]</mark>
Sodium N-Lauroylsarcosinate Hydrate	5g/25g <mark>[S0597</mark> ]

### **Amphoteric Surfactants**

Lauryl Sulfobetaine	5g / 25g [ <b>D3860</b> ]
Palmityl Sulfobetaine	5g / 25g <b>[H1283]</b>
Myristyl Sulfobetaine	5g/25g [ <b>T2653</b> ]
Caprylyl Sulfobetaine	5g / 25g [ <b>D4246</b> ]
n-Octyl Sulfobetaine	5g [ <b>D4247</b> ]

### **Nonionic Surfactants**

TRITON™ X-100 (n=approx. 10)	5g/25g [ <b>P1775</b> ]
Polyethylene Glycol Monocetyl Ether (n=approx. 23)	5g / 25g [P1776]
Polyethylene Glycol Monododecyl Ether (n=approx. 25)	5g / 25g [P1777]
Tween 20 (=Polyoxyethylene Sorbitan Monolaurate)	5g/25g <b>[T2530]</b>
Tween 40 (=Polyoxyethylene Sorbitan Monopalmitate)	5g/25g [ <b>T2531</b> ]
Tween 60 (=Polyoxyethylene Sorbitan Monostearate)	5g / 25g [ <b>T2532</b> ]
Tween 80 (=Polyoxyethylene Sorbitan Monooleate)	5g / 25g [ <b>T2533</b> ]
Tween 85 (=Polyoxyethylene Sorbitan Trioleate)	5g / 25g [ <b>T2534</b> ]
<i>n</i> -Octyl-β-D-Glucopyranoside	1g [ <b>O0355</b> ]

### Non-Detergent Sulfobetaines (NDSB)

Non-detergent sulfobetaines (NDSB) are amphiphilic small compounds containing both a cationic and anionic component which do not form micelles because of their small hydrophobic moiety. NDSBs solubilize proteins under mild conditions and can prevent protein denaturation by heat or acid, inhibit protein aggregation, acceleration protein refolding, and aid membrane protein extraction.

NDSB 211	1g/5g [ <b>H1399</b> ]
NDSB 201	5g/25g <mark>[S0813]</mark>
NDSB 256-4T	1g [ <b>B4030</b> ]

### Protease Inhibitors

Proteolysis is one of the major problems during protein extraction as they result in decreased yields. The addition of inhibitors helps prevent proteolysis and improves recovery of the desired protein. Inhibitors are also used during immunoprecipitation to prevent decomposition of antigens or antibodies by proteolytic impurities.

### **Cysteine Protease Inhibitors**

**2-lodoacetamide** 5g [10741] **E-64d** 5mg / 25mg [**E1337**]

### **Serine Protease Inhibitors**

**AEBSF** (=4-(2-Aminoethyl)benzenesulfonyl Fluoride Hydrochloride)

Benzamidine Hydrochloride 5g [B3379]
Benzylsulfonyl Fluoride 5g/25g [B3473]

### **Metalloprotease Inhibitors**

EDTA 2Na Dihydrate 5g/25g [D3789]
EDTA 3Na Hydrate 5g/25g [T2599]
EGTA 5g/25g [E0805]
1,10-Phenanthroline Monohydrate 5g [P1826]

### Protein Denaturation Reagents

Proteins fold into higher-order structures due to interactions such as hydrogen bonding, ionic interactions, and Van der Waals forces. Heat, acids and alkalis can change protein conformation and denature proteins. Protein extraction and analysis require protein denaturation, necessitating the use of urea and guanidine, which are chaotropic agents that disrupt the hydrogen bonding network.

 Guanidine Hydrochloride
 25g / 100g / 500g [G0197]

 Guanidine Thiocyanate
 5g / 25g [G0360]

 Thiourea
 5g / 25g [T2835]

 Urea
 5g / 25g [U0077]

## Nucleic Acid Removing Agents for Protein Sample Clarification

The process of nucleic acid removal may be effective in the purification of proteins. That is because the nucleic acid exhibits viscosity and the protein and the nucleic acid are likely to form a complex. The way to remove nucleic acid is absorption the nucleic acid to a basic water-soluble polymer or separation by binding and precipitating nucleic acid with nucleic acid removing agent such as streptomycin sulfate.

 Polyethyleneimine (ca. 30% in Water)
 25g / 100g [P1921]

 Streptomycin Sulfate
 5g / 25g [S0834]

### Preservatives and Disinfectants

Research in the life sciences requires the analysis of biological samples. Microorganisms can easily grow in these samples, and also in the buffers and reagents used for biological analysis. Therefore, preservatives are frequently added to samples and buffers to prevent the growth of microorganisms.

Amprolium Hydrochloride	5g/25g [A2572]
2-n-Octyl-4-isothiazolin-3-one	1g [ <b>O0378</b> ]
Dimetridazole	5g / 25g [D4081]
2-Chloroacetamide	5g / 25g [C2536]
5-Bromo-5-nitro-1,3-dioxane	5g [ <b>B3769</b> ]
1,2-Benzisothiazol-3(2 <i>H</i> )-one	5g [ <b>B3767</b> ]
Sorbic Acid Potassium Salt	5g / 25g [P1954]
Sorbic Acid	5g / 25g <b>[S0856]</b>
1,3-Butanediol	5g / 25g [B3770]
2-Phenoxyethanol	5g / 25g [P1953]
2-Hydroxybenzoic Acid	5g / 25g [H1342]
Benzoic Acid Sodium Salt	5g / 25g <b>[S0855]</b>
Benzylparaben	5g / 25g [B3768]
Isobutylparaben	5g / 25g [ <b>10816</b> ]
Butylparaben	5g / 25g [ <b>B3771</b> ]
Isopropylparaben	5g / 25g [ <b>10817</b> ]
Propylparaben	5g / 25g [P1955]
Ethylparaben	5g / 25g <b>[E0884]</b>
Methylparaben	5g / 25g [M2206]

### Protein Determination Reagents

The determination of protein concentration is essential for biochemical research. The following two products are supplied as a ready-to-use solution for quantitative protein determination.

### Pyrogallol red-molybdate protein assay

### **Pyrogallol Red** (Ready-to-use Solution) [for Protein determination]

100mL [P2575]

This product is supplied as a ready-to-use solution for protein determination based on the pyrogallol red-molybdate complex. When the dye binds proteins, the absorption maximum of the dye shifts from 480 nm to 600 nm in a linear manner with an increase in the quantity of the protein. It stains cuvettes very little, thus it can be washed with water alone after use.

### **Application**

- Prepare standard protein solutions with a series of dilutions.
- Mix P2575 with unknown protein samples, standard protein solutions and distilled water according to Table 1.
- 3) Incubate for 30 minutes at room temperature.
- 4) Measure absorbance at 600 nm.
- 5) Prepare a standard curve by plotting the absorbance data measured in #4) after subtracting from blank absorbance (distilled water), and calculate the amount of protein in test samples.

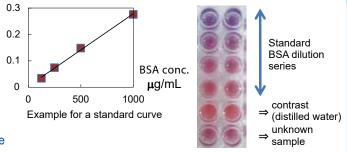
Table 1 : Volume for test tube or micro plate assay

Assay	test tube	micro plate
Measurement range	0.1 -1.0 mg/mL	0.1 -1.0 mg/mL
Sample solution or *protein standard	50 μL	10 μL
P2575	1 mL	200 μL

<sup>\*</sup>This product requires the standard protein solution (such as BSA).

### **Example for use: in a microplate**

- Prepare four dilution series of standard protein solutions from the concentration at 1000 mg/mL by doubling dilution.
- Mix 200 μL of P2575 with 10 μL each of a protein sample at an unknown concentration, the standard protein solution and distilled water in a 96 microplate.
- 3) Incubate for 30 minutes at room temperature, measure absorbance at 600 nm, and prepare a standard curve.



Example for a reaction

#### Compatible substance concentrations in protein sample of P2575

Substances at the following concentrations in the sample solutions do not affect the reaction results.

Buffers		Chelating Agents		Solvents	
Substance	Conc.	Substance	Conc.	Substance	Conc.
Glycine	100 mM	EDTA	100 mM	Acetone	10 %
Tris	2 M	EGTA	10 mM	DMSO	10 %
HCI	200 mM	Sodium citrate	200 mM	Ethanol	10 %
HEPES	100 mM			Methanol	10 %
MES	100 mM	Salts		Glycerol	10 %
MOPS	100 mM	Substance	Conc.		
PIPES	100 mM	(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	1 M	Denaturants	
Tricine	100 mM	KCI	1 M	Substance	Conc.
Imidazole	200 mM	MgCl <sub>2</sub>	50 mM	DTT	100 mM
Glucose	1 M	CaCl <sub>2</sub>	10 mM	Glutathione	1 mg / mL
Sucrose	25 %	NiCl <sub>2</sub>	10 mM	2-Mercaptoethanol	1 M
Fructose	1 M	ZnCl <sub>2</sub>	10 mM	Guanidine-HCI	1 M
		NaCl	2 M	Urea	3 M
		NaOH	100 mM		
		NaH <sub>2</sub> PO <sub>4</sub>	500 mM	Detergents	
		NaN <sub>3</sub>	0.50 %	Substance	Conc.
		· · ·		SDS	0.10 %
				Triton X-100	0.10 %
				Tween-20	0.10 %

### **Bradford assay**

### **Bradford Assay Solution** (Ready-to-use Solution) [for Protein determination] 500mL [B5702]

This product is supplied as a ready-to-use solution for protein assay based on the method of Bradford. This product contains Coomassie Brilliant Blue G-250 (CBB G-250). When the dye containing CBB G-250 binds proteins, the absorption maximum of the dye shifts from 465 to 595 nm linearly with the quantity of the protein. Absorbance can be measured only 5 minutes after the reaction starts. Low concentration of protein  $(1.0 - 25 \,\mu\text{g/mL})$  can be measured.

### **Application**

- 1) Prepare standard protein solutions with a series of dilutions.
- 2) Mix **B5702** with unknown protein samples, standard protein solutions and distilled water according to Table 2.
- 3) Incubate for 5 minutes at room temperature.
- 4) Measure absorbance at 600 nm.
- 5) Prepare a standard curve by plotting the absorbance data measured in #4) after subtracting from blank absorbance (distilled water), and calculate the amount of protein in test samples.

Table 1: Volume for test tube or micro plate assay

Assay	test tube	micro plate	micro assay
Measurement range	0.1 -1.0 mg/mL	0.1 -1.0 mg/mL	0.1 - 25 μg/mL
Sample solution or *protein standard	20 μL	4 μL	500 μL
B5702	1 mL	200 μL	500 μL

<sup>\*</sup>This product requires the standard protein solution (such as BSA).

#### Compatible substance concentrations in protein sample of B5702

Substances at the following concentrations in the sample solutions do not affect the reaction results.

Buffers		Salts		Solvents		Denaturan	ts
Substance	Conc.	Substance	Conc.	Substance	Conc.	Substance	Conc.
Glycine	100 mM	(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	1 M	Acetone	10 %	DTT	100 mM
Tris	2 M	KCI	1 M	DMSO	10 %	Glutathione	1 mg/mL
HCI	100 mM	MgCl <sub>2</sub>	50 mM	Ethanol	10 %	2-Mercaptoethanol	1 M
HEPES	100 mM	CaCl <sub>2</sub>	10 mM	Methanol	10 %	Guanidine-HCI	1 M
MES	100 mM	NiCl <sub>2</sub>	10 mM	Glycerol	10 %	Urea	ЗМ
MOPS	100 mM	ZnCl <sub>2</sub>	10 mM				
PIPES	100 mM	NaCl	2 M	Detergents		Chelating Ag	gents
Glucose	1 M	NaOH	100 mM	Substance	Conc.	Substance	Conc.
Sucrose	25 %	NaH <sub>2</sub> PO <sub>4</sub>	500 mM	SDS	0.05 %	EDTA	100 mM
Fructose	1 M	NaN <sub>3</sub>	0.50 %	Triton X-100	0.10 %	EGTA	10 mM
				Tween-20	0.10 %	Sodium citrate	200 mM

These products (P2575, B5702) require the protein standard solution T3796.

### **Related Product**

New Standard Solution of Albumin from Bovine Serum

5mL [T3796]

### **Bicinchoninic Acid (BCA) assay**

**New Bicinchoninic Acid Disodium Salt** [for Protein Research]

5g [**B5838**]

### Electrophoresis Reagents

Electrophoresis is a technique which separates charged biomolecules based on the rate at which they migrate in an applied electrical field. The following products are used in the Laemmli method, reagents widely used in protein staining and other related reagents.

### Reagents for gel preparation, buffer preparation, etc.

New 2X SDS-PAGE Sample Buffer (2-Mercaptoethanol free)	25mL [B5834]
New 4X SDS-PAGE Sample Buffer (2-Mercaptoethanol free)	20mL [B6140]
New 30% Acrylamide / Bis-acrylamide (29:1)	250mL [A3217]
New 30% Acrylamide / Bis-acrylamide (37.5:1)	250mL [A3218]
Acrylamide Monomer	25g / 500g [A1132]
Ammonium Peroxodisulfate	5g / 25g [ <b>A2098</b> ]
Bromophenol Blue Sodium Salt (= BPB)	1g [ <b>B3195</b> ]
DL-Dithiothreitol (= DL-DTT)	1g / 5g [D3647]
Glycerol	1g <b>[G0316]</b>
Glycine	25g / 500g [G0317]
2-Mercaptoethanol	5g / 25g [M1948]
<i>N,N</i> '-Methylenebisacrylamide	25g / 100g [M0506]
Sodium Dodecyl Sulfate (= SDS)	25g / 500g [ <b>S0588</b> ]
N,N,N',N'-Tetramethylethylenediamine (= TEMED)	5g / 25g <b>[T2515</b> ]
Tris(hydroxymethyl)aminomethane (= Tris-Base)	25g / 500g [ <b>T2516</b> ]

### **Protein Staining Reagent**

New Coomassie Brilliant Blue G-250 (Ready-to-use solution) [for Electrophoresis]

500mL [C3488]

### **Application**

- 1) After electrophoresis, wash the gel with deionized water for 5 minutes three times.
- 2) Remove the water used for washing, add C3488 till the gel is soaked, and let the gel stain for 1 hour while shaking gently at room temperature.
- 3) Remove the staining solution, destain the gel with deionized water for 1 hour and check it.
- 4) If the background is high, destain the gel with deionized water overnight at room temperature.

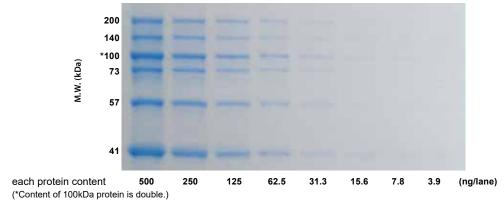


Figure. Proteins stained by the above method (destained overnight)

### Reagents for protein staining and others

Acid Black 1 (= Amido Black 10B)	5g <b>[A2097</b> ]
Acid Red 112 (= Ponceau S)	1g/5g <mark>[A2256]</mark>
Coomassie Brilliant Blue G-250	5g [ <b>B3193</b> ]
Coomassie Brilliant Blue R-250	5g [ <b>B3194</b> ]
Fast Green FCF	5g <b>[F0718</b> ]
Sodium Deoxycholate	25g [ <b>D1820</b> ]
6-Aminohexanoic Acid	5g / 25g [A2255]

### Nucleic Acid Detecting Reagents

### **Nucleic Acid Staining Reagent**

**New Ethidium Bromide** (0.5mg/mL in Water) (in Dropper Bottle) [for Electrophoresis] 10mL [E1363]



Each drop contains 20  $\mu$ g of Ethidium Bromide, so you can easily adjust the solution as final concentration. Convenient and safe to use because of dropper bottle.

### **Application**

After electrophoresis, dilute <code>E1363</code> (1 drop / 40 mL) to 0.5  $\mu g/mL$  with water or running buffer and incubate the gel for 15 min. If you have to decrease background fluorescence, wash the gel in water for 15 minutes.In use of electrophoresis buffer solution, Ethidium Bromide incorporated into nucleic acid and can visualize band immediately by using UV transilluminator.

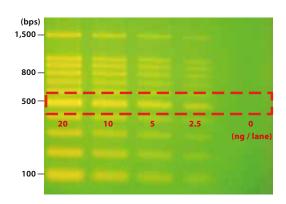


Figure. DNA Ladder Marker stained by the above method (destained 15 min)

# Protein-maleimide Conjugates for Thiol-maleimide Crosslinking

Bovine Serum Albumin Maleimide Conjugate (1mg×3)

1set [B5944]

Horseradish Peroxidase Maleimide Conjugate (0.5mg×3)

1set [H1621]

Streptavidin Maleimide Conjugate (0.5mg×1)

1vial [T3531]

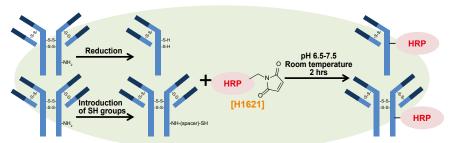
### Advantages

- Each product containing a thiol-reactive maleimide group can be used for the conjugation to proteins and peptides containing free thiols.
- Each protein conjugate is packaged for single use purposes and thus does not require weighing prior to use.

### Application: HRP-labelling of an antibody with H1621

In case of antibodies without free thiol (SH, sulfhydryl) groups, disulfide moieties in proteins can be reduced by a reductant such as DTT [D3647] or 2-MEA [A0296] to reveal free thiols.

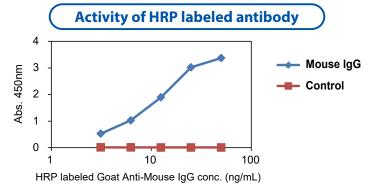
Furthermore, thiol group can be introduced to primary amines by adding SATA [S0431], SATP [S0859] or 2-Iminothiolane.



Example protocol for antibody conjugation starts from a reduction of native disulfide bonds in the Goat Anti-Mouse IgG, followed by labeling with the HRP using H1621. For more information, see the product detail page of H1621 on TCI website.

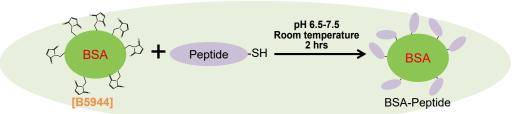
#### Protocol

- 1) Add DTT to a final concentration equal to 3 mole equivalents per mole equivalent of antibody present.
- 2) Incubate for 90 minutes at 37 °C.
- 3) Purify the reduced IgG by gel filtration or ultrafiltration, dialysis.
- 4) Add equal amount of H1621 (by weight) to a purified antibody and Incubate for 2 hours at room temperature (25 °C).



Goat Anti-Mouse IgG labeled with the HRP using H1621 was tested by ELISA for detection of a Mouse IgG coated on a plate. Mouse IgG could be detected sufficiently even if the labeled antibody was diluted to 5 ng/mL or more.

### Application: Preparation of BSA-Peptide using **B5944**



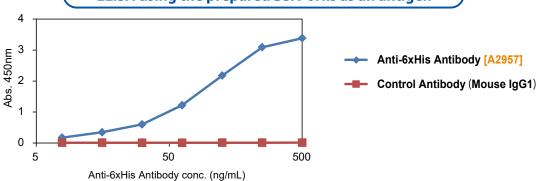
BSA is conjugated to haptens and typically used as an antigen carrier for anti-hapten antibody. Here we show how to conjugate 6xHis-Cys peptide to B5944.

For more information, see the product detail page of B5944 on TCI website.

#### **Protocol**

- 1) Dissolve the 6xHis-Cys peptide in 0.1 M sodium phosphate, 0.15 M NaCl, 0.1 M EDTA at pH 7.2.
- 2) Reconstitute the B5944 with 100 μL of water.
- 3) Add 1mg of 6xHis-Cys peptide to 1 mg of B5944 and Incubate for 2 hours at room temperature (25 °C).

### ELISA using the prepared BSA-6His as an antigen



Anti-6xHis Antibody [A2957] was analysed by ELISA using a 0.1  $\mu g$  / well of BSA-6His coated plate.

Goat Anti-Mouse IgG HRP Conjugate [G0407] was used as the secondary antibody.

### **Related Products**

#### **Reducing agents for protein disulfide**

DTT (= DL-Dithiothreitol)

2-MEA (= 2-Aminoethanethiol Hydrochloride)

2-Mercaptoethanol

Tris(2-carboxyethyl)phosphine Hydrochloride

**Reagents for introduction of thiol group** 

SATA (= N-Succinimidyl S-Acetylthioglycolate)

SATP (= *N*-Succinimidyl 3-(Acetylthio)propionate)

1g / 5g [**D3647**]

25g / 100g / 500g [A0296]

5g / 25g [M1948]

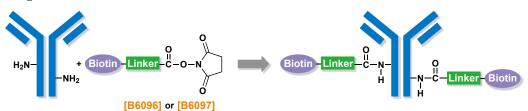
1g / 5g / 25g [T1656]

1g / 5g [S0431] 100mg [S0859]

### Pre-Weighed Biotinylation Reagents

**Biotin-LC-LC-NHS** (2mg×5) **Biotin-PEG<sub>2</sub>-NHS** (2mg×5)

1set [**B6096**] 1set [**B6097**]



### **Advantages**

B6096 and B6097 contain both a linker and an N-hydroxysuccinimidyl ester moiety, and easily react with amino group (-NH $_2$ ) of proteins. Target samples can be biotinylated without weighing of the products during the preparation. B6096 and B6097 include 5 reagent vials, each containing 2 mg of respective reagent. The pre-aliquoted packaging prevents decline of the reagent reactivity over time by eliminating the need for repetitive opening of the vial.

### **Applications**

### Preparation:

Use of a 10 mM biotinylation solution is recommended. In order to efficiently biotinylate a sample, biotinylation solution should be used at a 15-fold molar excess over the amount of amine-containing protein. Make sure to calculate the 10 mM biotinylation solution amount (see example below).

Calculate: A µL of 10 mM biotinylation solution for biotinylation of 2 mg lgG (150,000 M.W.)

2 [mg lgG] x 10<sup>-3</sup> [g/mg] x 1/150,000 [mol/g] x 15 [fold]

= A [ $\mu$ L of 10 mM biotinylation solution] x 10<sup>-6</sup> [L/ $\mu$ L] x 10 [mmol/L] x 10<sup>-3</sup> [mol/mmol]

A = 20 [ $\mu$ L of 10 mM biotinylation solution]

#### **Direction for Use:**

- 1. Bring each product to room temperature.
- 2. Dissolve 2 mg of Biotin-LC-LC-NHS [B6096] in 350  $\mu$ L of DMSO or DMF or 2 mg of Biotin-PEG<sub>2</sub>-NHS [B6097] in 400  $\mu$ L of PBS to prepare a 10 mM biotinylation solution.
- 3. Dissolve the sample (1-10 mg/mL) in an appropriate buffer such as PBS. Do not use buffers including amines (such as Tris).
- 4. Add A  $\mu$ L of 10 mM biotinylation solution to the sample solution and incubate the mixed solution for 30 min at room temperature.
- 5. Remove unreacted and hydrolyzed reagent using desalting column or dialysis methods.

### **Related Products**

Biotin-LC-LC-NHS	25mg / 100mg [S0956]
Biotin-PEG,-NHS	25mg / 100mg [S0955]
Biotin-PEG,-Maleimide	50mg [B3174]
Streptavidin from Streptomyces avidinii	1mg/vial [S0951]
Streptavidin HRP Conjugate	0.1mg/vial [S0972]
Streptavidin FITC Conjugate	0.1mg/vial [S0966]
Streptavidin Maleimide Conjugate	0.5mg/vial [ <b>T3531</b> ]
НАВА	5g / 25g [H0586]
Sulfo-SMCC Sodium Salt	20mg / 100mg [S0883]
Horseradish Peroxidase Maleimide Conjugate (0.5mg×3)	1set [H1621]
BSA Maleimide Conjugate (1mg×3)	1set [B5944]

### **Ordering and Customer Service**

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