

# jetOPTIMUS<sup>®</sup> transfection reagent

## Short protocol - DNA Transfection

### DAY 0: Cell seeding

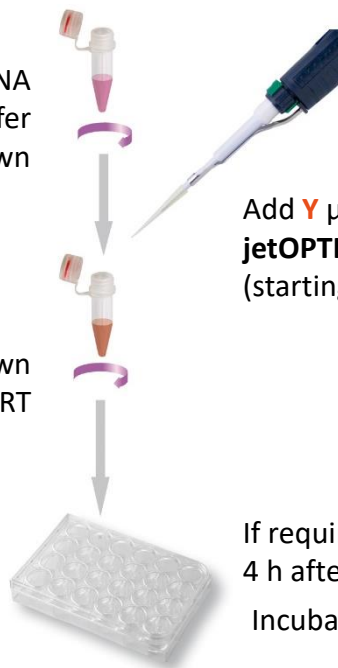
→ Seed cells in **V** ml of serum containing medium according to the table below

#### Quantities per well, dish or flask

Culture vessel	Number of cells	V = volume of serum containing medium during transfection
96-well	7500 - 25 000	0.125 mL
24-well	40 000 - 100 000	0.5 mL
6-well / 35 mm	150 000 - 400 000	2 mL
60 mm / flask 25 cm <sup>2</sup>	200 000 - 850 000	5 mL
100 mm / flask 75 cm <sup>2</sup>	1 x 10 <sup>6</sup> - 4 x 10 <sup>6</sup>	10 mL

### DAY 1: Transfection using jetOPTIMUS<sup>®</sup> reagent

Dilute **X** µg of DNA  
in **W** µl of jetOPTIMUS<sup>®</sup> buffer  
Vortex 1 second and spin down



- Transfect cells at **60-80% confluency**
- Use **jetOPTIMUS<sup>®</sup> buffer only**
- Perform transfection **in the presence of serum**

Add **Y** µl of  
**jetOPTIMUS<sup>®</sup> reagent**  
(starting ratio 1:1)

Vortex 1 second, spin down  
and incubate 10 min at RT

Add transfection mix to cells in  
serum-containing medium

If required, replace medium  
4 h after transfection  
Incubate 24 to 48 h

#### Quantities per well, dish or flask

Culture vessel	W = volume of jetOPTIMUS <sup>®</sup> buffer	X = amount of DNA added	Y = volume of jetOPTIMUS <sup>®</sup> reagent
96-well	12.5 µL	0.13 µg	0.13 – 0.19 µL
24-well	50 µL	0.5 µg	0.5 – 0.75 µL
6-well / 35 mm	200 µL	2 µg	2 – 3 µL
60 mm / flask 25 cm <sup>2</sup>	500 µL	4 µg	4 – 6 µL
100 mm / flask 75 cm <sup>2</sup>	1000 µL	10 µg	10 – 15 µL

### DAY 2-3: Measure gene expression

See back page for optimization tips

Download complete protocol on [www.polyplus-transfection.com/resources/product-literature/](http://www.polyplus-transfection.com/resources/product-literature/)

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## Short protocol - Optimization Tips

### + Protocol Optimization

- + Test different DNA amounts: X, 0.5 X and 1.5 X
- + Test different DNA/jetOPTIMUS<sup>®</sup> ratios, 1:1 to 1:1.5

#### Quantities per well, dish or flask

Culture vessel	W = volume of jetOPTIMUS <sup>®</sup> buffer	X = amount of DNA added	Y = volume of jetOPTIMUS <sup>®</sup> reagent
96-well	12.5 µL	0.10 – 0.20 µg	0.10 – 0.30 µL
24-well	50 µL	0.25 – 0.75 µg	0.25 – 1 µL
6-well / 35 mm	200 µL	1 – 3 µg	1 – 4.5 µL
60 mm / flask 25 cm <sup>2</sup>	500 µL	2 – 6 µg	2 – 9 µL
100 mm / flask 75 cm <sup>2</sup>	1000 µL	5 – 15 µg	5 – 22 µL

- + Check our online Transfection Database for cell specific protocols at:  
[www.polyplus-transfection.com/resources/cell-transfection-database/](http://www.polyplus-transfection.com/resources/cell-transfection-database/)

### + Tips to increase cell viability of sensitive cells

- + Replace medium after 4 h.
- + Decrease DNA amount to 0.5 X while maintaining the DNA/jetOPTIMUS<sup>®</sup> ratio previously used.
- + Analyze transfection at an earlier time point (24 h after transfection instead of 48 h for instance).
- + Check that the target gene does not affect cell viability.
- + Ensure that cells have been passaged more than twice and less than 20 times prior to transfection. Discard overconfluent cells.

### + Good DNA Transfection Practices

- + Store appropriately jetOPTIMUS<sup>®</sup> (5 ± 3°C).
- + Regularly check for mycoplasma contamination.
- + Use a reporter gene to set up and optimize transfection conditions.
- + Serum quality may drastically affect transfection efficiency. When purchasing a new batch of serum or trypsin, check cell viability as well as transfection efficiency.

**Note:** Please refer to the complete protocol available online at:  
[www.polyplus-transfection.com/resources/product-literature/](http://www.polyplus-transfection.com/resources/product-literature/)