

Data sheet

GreenSafe DNA gel stain

Cat. No: E0206

Cat. No: E0207

Introduction

GreenSafe DNA gel stain product replace toxic Ethidium Bromide (EtBr) for the visualization of double-stranded DNA, single-stranded DNA, and RNA in agarose and polyacrylamide gel electrophoresis.

GreenSafe DNA gel stain has 2 fluorescence excitation wavelengths in the UV range (~270nm; ~290nm) and one in the blue light range (~485nm). Maximum fluorescence emission is at ~525nm (green). Therefore, **GreenSafe DNA gel stain** is compatible with a large variety of gel documentation systems.

Features

- **Easy to Use:** you can directly replace EB with without changing your existing imaging system.
- **Safe:** Non-carcinogenic by the AMES test.
- **Sensitive:** Increase your sensitivity by reducing nonspecific background fluorescence.

Applications

Non-carcinogenic alternative to Ethidium bromide.

Storage

You may store the **GreenSafe DNA gel stain** at any temperature between 2°C to 8°C protected from light. Do not freeze!

PRODUCT USE LIMITATION

This product is developed, designed and sold exclusively for research purposes and in vitro use only. The product was not tested for use in diagnostics or for drug development, and is not suitable for administration to humans or animals. Please refer to www.canvaxbiotech.com for the Material Safety Data Sheet of the product.

Kit Contents

	E0206	E0207
GreenSafe DNA gel stain	500 µl	1 mL

Pre-casting protocol:

500 µl of this stain is sufficient for 10 L of agarose gel.

1. Prepare 100 ml of agarose gel solution (concentration from 0.8 - 3.0 %) and heat until the solution is completely clear and no small floating particles are visible.
2. Add 4-6 µl of the stain to the gel solution and mix it gently.
3. Cool the gel to 50 – 60 °C and cast the gel into the gel tray.
4. When the gel is solid, load the samples and perform electrophoresis.
5. Detect the bands under UV illuminator. Can also be viewed with Blue LED light. Yellow or green gelatin or cellophane filters should be used for photography.

Poststaining

1. For <0.5 cm thick agarose gels, 10-15 µL of stain should be used per 100 mL of buffer
2. Incubate the gel in staining solution for 10 - 30 minutes. Staining time varies with the thickness of the gel and percentage of the agarose.
3. The post-staining solution may be used 2-3 times. Staining solution that is going to be reused should be preferably stored at room temperature in the dark.

Notes

